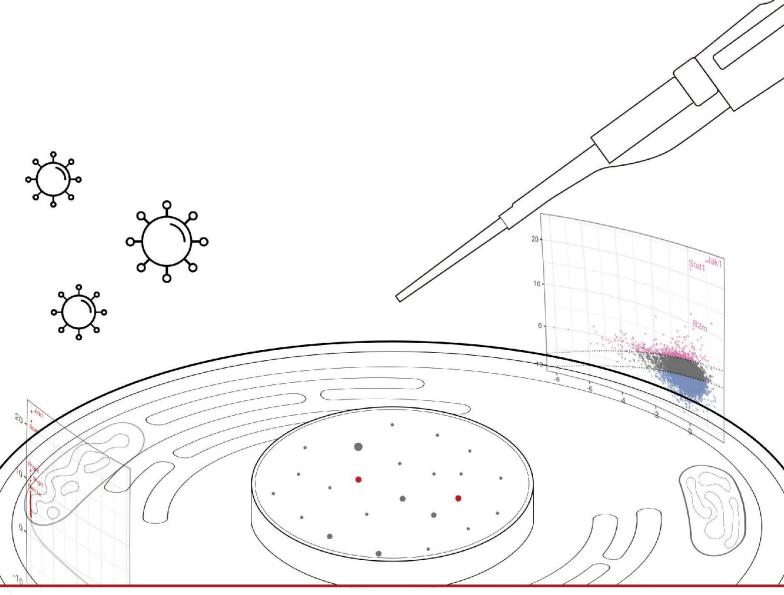
# **CRISPR Library**

Instructions



TEL.: +86 153 6067 3248 (Intl)

Web: www.ubigene.us

Add.: Floor 12, Building 2A, GZ-BJ Innovation Center, No. 45, Ruiji 2nd Street, Huangpu District, Guangzhou, 510000, P.R.China

US Toll free: 855 777 3210

EU& APAC Toll free: 800 3272 9252

Mail: info@ubigene.com



# Use Instruction - Human GeCKO v2 genome-wide plasmid library (Single-plasmid system)

#### Product Info

This Human GeCKO v2 genome-wide plasmid library (Single-plasmid system) is applicable for human genome-wide knockout and gene screening, containing over 100,000 knockout vectors targeting all exons of the whole genome. It consists of two half-libraries, library A and library B, each half-library contains 3 gRNAs per gene. So libraries A and B together contain 6 gRNAs per gene. Libraries A and B can be used in combination or alone as per requirement. The vector backbone of the library is LentiCRISPRv2 backbone which is the all-in-one vector system, that is, the gRNA and Cas9 gene are on the same vector.

# Library Details

Product Name	Human GeCKO v2 genome-wide plasmid library		
	(Single-plasmid system)		
Product Catalog	YKO-Libr-H001A, YKO-Libr-H001B		
Product Details	123,411 gRNAs (gRNA sequences see attachment);		
	Single-plasmid system, can be directly used for library screening without first constructing Cas9 stable cell lines;		
	Puromycin resistance, puromycin can be used for antibiotic screening upon cell infection;		
	Plasmids paired with 3 <sup>rd</sup> lentivirus packaging system can be		

+86 153 6067 3248 | US Toll free: 855 777 3210 | EU Toll free: 800 3272 9252 | Korea Toll free: 001 800 3272 9252

	directly used for virus packaging.  *It is recommended to use Ubigene's Lentiviral Packaging Kit (Cat# YK-LVP-05)			
	Library A 65,383 gRNAs;			
		Targeting 19,050 genes, 3 gRNAs per gene;		
		Targeting 1,864 miRNAs, 4 gRNAs per miRN		
		1,000 non-target control sgRNAs		
	Library B	58,028 gRNAs;		
		Targeting 19,050 genes, 3 gRNAs per gene;		
		1,000 non-target control sgRNAs		
Backbone Map	cPPT psi+ RRE U6 sgRNA EFS SpCas9 FLAG P2A Puro WPRE			
Verification Primers	LentiCRISPRv2-F: ATTTCTTGGGTAGTTTGCAGTTT			
	LentiCRISPRv2-R: GACTCGGTGCCACTTTTTCA			
Product Specifications	Ready-to-use, endotoxin-free, maxiprep plasmids, verified			
	by Next Generation Sequencing, with coverage>99% and			
	uniformity<10.			

# **■ Product Use Instruction**

# Part 1. Lentivirus Packaging

Mix library plasmid constructs with 3<sup>rd</sup> generation lentiviral packaging constructs to be co-transfected into 293T cells (Recommend: Ubigene's 293T cell line specialized for virus packaging, cat#YC-A006). 48 or 72 hours upon transfection, collect lentiviral

supernatant and the virus can be used upon concentration. The virus should be stored at -80°C.

#### **Part 2. Library Plasmid Amplification**

1. Library plasmid electroporation

Add 100 ng library plasmid to 25  $\mu$ L electrocompetent cells with transformation efficiency  $\geq 10^9$  cfu/ug, electroporate cells as per electroporation parameters. Upon electroporation, add 975  $\mu$ L recovery medium, mix well and transfer to a tube, then add 1 ml recovery medium to the tube and mix well again. Repeat above steps for three times and get 4 electroporation end products, share the tubes at 37°C, 250 rpm for 1 hour.

- 2. Culture of amplified library and calculation of transformation efficiency
- 1) Mix the 4 tubes of electroporation end products and take 10  $\mu$ L and dilute with 990  $\mu$ L recovery medium. Plate 20  $\mu$ L dilution onto a 10 cm Petri dish and incubate plates at 32°C for 14 hours. Count the colonies in the dish. If the number of colonies is 40000X greater than 6 x 10<sup>6</sup>, move on to the next step. If it is less than 6 x 10<sup>6</sup>, redo this step.
- \* Note: It is recommended that the number of colonies should be 40000X greater than  $2 \times 10^7$  to ensure the uniformity of Library gRNA
- 2) For the remaining electroporation end products, plate 400  $\mu$ L/dish (20 dishes can be plated in total), then incubate at 32°C for 14 hours.
- 3. Collect transformation products
  - 1) Add 500 µL LB medium to each dish, use spreader to scrape the plates and

collect the bacteria to a 50 mL centrifuge tube

- 2) Repeat the steps for all plates
- 3) Centrifuge tubes to pellet bacteria, decant LB and weigh pellet (bacteria).

#### 4. Maxiprep

Maxiprep the plasmid DNA according to the instruction for maxiprep kit, it is recommended to use endofree maxiprep kit from QIAGEN, MACHEREY-NAGEL, etc (e.g. EndoFree Plasmid Mega Kit from QIAGEN)

#### Part 3. Library Screen

#### 1. Determine infect MOI

Dilute the library virus into different gradients, such as MOI=0.3, 0.5, 1, 5, 10, 30, 100 to infect the target cells (the cell confluency is 30-50%). Each gradient needs to be set with 2 wells. After 48 hours of infection, add puromycin according to the settings in the table below for screening, and stop antibiotic screening when all cells in the blank group (cells not infected with virus) die. The MOI with a survival rate of 30% after antibiotic screening is the virus infection condition for the library screening experiment, that is, infect MOI (MOI=0.3 in some literatures actually refers to the amount of virus corresponding to the virus infection of 30% cells).

Group#	MOI	Antibiotic	Cell amount	Survival rate upon
		screening	upon antibiotic	antibiotic screening
			screening	
Experimental	0.3	Yes	N1	N1/M1
group 1				
Experimental	0.5	Yes	N2	N2/M2
group 2				
Experimental	1	Yes	N3	N3/M3
group 3				
Experimental	5	Yes	N4	N4/M4



group 4				
Experimental	10	Yes	N5	N5/M5
group 5				
Experimental	30	Yes	N6	N6/M6
group 6				
Experimental	100	Yes	N7	N7/M7
group 7				
Infection blank	0.3	No	M1	
group 1				
Infection blank	0.5	No	M2	
group 2				
Infection blank	1	No	M3	
group 3				
Infection blank	5	No	M4	
group 4				
Infection blank	10	No	M5	
group 5				
Infection blank	30	No	M6	
group 6				
Infection blank	100	No	M7	
group 7				
Blank group	0	Yes		

# 2. Transduction of target cells with library virus

1 Determine the amount of cells and virus

Cell amount= 
$$\frac{gRNA\# \times gRNA \text{ coverage}}{30\%}$$
 \* gRNA coverage>500 fold

Virus amount = cell amount × infect MOI

- ② Expand the cells according to the cell amount calculated in step ①, and prepare sufficient virus.
- ③ Use library virus infect the target cells, upon puromycin screening, divide the screened cells into experimental group and control group. Add target drugs to the experimental group for screening, upon screening, collect 5 x 10<sup>7</sup> cells and perform genome extraction for Next Generation Sequencing, and then compare and analyze the gRNAs of the experimental group and the control group.



# Relevant products and service

Ubigene provides 35+ off-shelf libraries including Human/Mouse genome-wide plasmid library and some sub-libraries, and one-stop customized screening services for CRISPR-KO, CRISPRa, and CRISPRi including high-throughput sgRNA library construction, virus packaging, cell infection, drug screening, NGS sequencing, and data analysis, etc. Multiple deliverables fulfill different research needs!